Association of the Treatment Induced Clearance of Hepatitis C Virus Infection with the IL28B Gene Polymorphisms

Case Report

Introduction

Hepatitis C virus (HCV) has often been referred to as the “silent virus,” as most HCV infections are clinically silent until the disease reaches a late stage, which often occurs several decades after initial infection. Although a variety of host factors play a role in eradication of HCV, only 15%-25% of adults spontaneously clear the infection. Although a variety of host factors play a role in eradication of HCV, only 15%-25% of adults spontaneously clear the infection. The remaining 75%-85% of patients continue to have persistent viremia, life-long HCV infection, chronic hepatitis C [2].

In the early 2000s, the combination of pegylated interferon alpha (PEGIFNα) and daily doses of ribavirin (RBV) became the standard-of-care (SOC) treatment for chronic hepatitis C [3,4]. The primary goal of the treatment is eradication of HCV, which is synonymous with sustained viral response (SVR), defined as undetectable HCV RNA in blood, 24 weeks after completion of the antiviral treatment [5]. The PEGIFN/RBV treatment is prolonged and expensive, complicated by side effects leading to treatment discontinuation, and only about one-half of the treated patients infected with HCV genotype 1 are achieving sustained viral response [6].

A novel family of antiviral cytokines was described and designated as type III interferons. The interferon type III family consists of three cytokines: Interleukin 29 (interferon lambda 1, IFNλ1), Interleukin 28A (interferon lambda 2, IFNλ2), and Interleukin 28B (interferon lambda 3, IFNλ3). IFNλ is rapidly induced during HCV infection and has antiviral activity against the virus. The type III interferons induce antiviral activity through both innate and adaptive immune pathways [7-9].

The IL28B gene encodes the cytokine interferon lambda 3, IFNλ3. Four genome wide association studies (GWAS) had associated treatment-induced clearance of HCV following PEGIFN/RBV therapy with the single nucleotide polymorphisms (SNPs) near the interleukin 28B (IL28B) gene on chromosome 19 [10-13]. Differences in genetic sequences among individuals are called genetic polymorphisms. The GWAS showed that SNPs near the IL28B gene (CC of rs12979860, TT genotype of rs8099917, and AA genotype of rs12980275) were significantly associated with sustained viral response (p=0.006, p=0.002, p=0.007, respectively) in study patients with chronic hepatitis C treated with standard antiviral therapy.

Aim: The aim of the study was to evaluate the association between the SNPs near the IL28B gene and the response to the treatment of chronic hepatitis C.

Methods: The genotyping of the three IL28B gene polymorphisms: rs12979860, rs8099917, and rs12980275 was done in 100 Caucasian patients with chronic hepatitis C previously treated with standard antiviral therapy. The study group consisted of 28 hemodialysis patients with end stage renal disease treated with pegylated interferon α and 72 patients without renal disease treated with pegylated interferon α and ribavirin. All patients finished the antiviral treatment at least 6 months before enrollment in the study. Sustained viral response, defined as an absence of detectable HCV RNA in the serum, was tested by an assay with a sensitivity of 20 IU/mL.

Results: Sustained viral response was achieved in 56% (56/100) of the treated patients. The three IL28B gene polymorphisms (CC genotype of rs12979860, TT genotype of rs8099917, and AA genotype of rs12980275) were associated with sustained viral response (p=0.006, p=0.002, p=0.007, respectively) in study patients with chronic hepatitis C treated with standard antiviral therapy.

Conclusion: The IL28B gene polymorphisms: rs12979860, rs8099917, and rs12980275 were significantly associated with the successful treatment of chronic hepatitis C.
The aim of the study was to evaluate the association between single nucleotide polymorphisms near the IL28B gene and the response to the treatment of chronic hepatitis C.

**Patients and Methods**

**Patients**

A study group of 100 adult Caucasian patients with chronic hepatitis C routinely treated with antiviral therapy was investigated in the study. The study was approved by the local Ethics Committee and written informed consent was obtained from each study participant. The cohort consisted of 28 patients with end stage renal disease on hemodialysis (HD) treatment and 72 patients without renal disease (non-renal). The HD patients received antiviral therapy only with pegylated interferon α-2a (PEGIFNa-2a). The non-renal patients were treated with pegylated interferon α-2a or α-2b (PEGIFNa-2b) plus ribavirin. The pegylated interferon was administered subcutaneously once a week in a standard dose (PEGIFNa-2a: 135 µg for HD patients and 180 µg for non-renal patients, and PEGIFNa-2b: 1.5µg/kg). The ribavirin was administered daily in a dose of 1000 mg for patients with body weight less than 75 kg and 1200 mg for patients with body weight over 75 kg when was combined with PEG IFNa-2a. The ribavirin dose was 800 mg for patients with body weight less than 65 kg, 1000 mg for patients with body weight 65–85 kg and 1200 mg for patients with body weight over 85 kg when was combined with PEG IFNa-2b. The treatment duration was 24 weeks for patients infected with HCV genotype 2 and 3, and 48 weeks for the infection caused by HCV genotype 1 and 4. All patients finished the antiviral treatment at least 6 months before enrollment in the study. Sustained viral response (SVR), defined as an absence of detectable HCV RNA in the serum, 6 months after completion of the antiviral treatment, was tested by an assay with a sensitivity of 20 IU/mL.

**Methods**

Peripheral blood samples in EDTA as an anticoagulant were obtained from each patient enrolled in the study.

**HCV quantification**

Hepatitis C virus RNA was extracted from plasma using QIAamp Viral RNA kit (Qiagen, Hilden, Germany) according to manufacturer’s instructions. Reverse transcriptase- polymerase chain reaction (RT-PCR) assay for HCV quantification was done with HCV Real-TM Quant (Sacace Biotechnologies, Como, Italy) on Stratagene MX3005P real-time PCR system (Agilent Technologies, Edinburgh, UK) according to manufacturer’s instructions. Detection limit of the assay is 20 IU/ml.

**IL28B polymorphisms genotyping**

Peripheral blood mononuclear cells (PBMCs) were isolated by density-gradient centrifugation using Histopaque-1077 (Sigma-Aldrich, Munich, Germany) and homogenized in Tri Reagent Solution (Ambion, Life Technologies and Carlsbad, CA, USA). Genomic DNA was extracted from PBMCs homogenized in Tri Reagent Solution (Ambion, Life Technologies, Carlsbad, CA, USA) according to manufacturer’s instructions and genotyped for three IL28B polymorphisms: rs8099917, rs12979860 and rs12980275. The rs8099917 polymorphism was genotyped using TaqMan predesigned SNP genotyping assay (reference C_11710096_10, Applied Biosystems) according manufacturer’s recommended protocol in a total volume of 25µl. The two later SNPs were genotyped using custom designed TaqMan assays with the following primers and probes: TCTACTGAAACAGGAGACCT, GCGGGGATGGCAATTTCAAC, 6Fam-TGGTTCCACGCTTC, Vici-TGGTTCCCGCTC for rs12979860, and GTGCTGAGAAGTCAAATTCC, CCGTACCCCGCAAAATT, 6Fam-ACACGTCGTTTCTTA, Vic-AGACACGTCGTCTTCTC for rs12980275, and rs12979860, GTGCTGAGAAGTCAAATTCC, CCGTACCCCGCAAAATT, 6Fam-ACACGTCGTTTCTTA, Vic-AGACACGTCGTCTTCTC for rs12980275. For both assays 20ng of DNA was used in a total volume of 25µl including 12.5µl TaqMan Universal PCR master Mix (2x), 1µM of each primer and 200nM of each probe. The PCR reaction conditions were as follows: initial denaturing at 95°C for 10min; 40 cycles of 15 sec at 92°C and 1 min at 64°C to reduce miss priming. Thermal cycling was performed using a Stratagene MX3005P real-time PCR system (Agilent Technologies, Edinburgh, UK). Both positive and negative controls were included in every genotyping assay.

**Statistical analysis**

Statistical analysis of data was performed using SPSS Statistics 17.0. The parametric variables are presented as the mean and standard deviation. A logistic regression model was used to determine the predictors of the SVR. Odds ratios (ORs) and 95% confidence intervals (95% CIs) were derived from the logistic regression model. The Fisher’s exact test was used to compare proportions. A p value was calculated with two tails. A two-tailed p value of less than or equal to 0.05 was considered statistically significant.

**Results**

The study group included 100 patients, 71 (71%) men and 29 (29%) women, with a mean age of 43.5 ± 11.4 years. All patients were Caucasians. The HCV genotype 1 caused the chronic hepatitis C infection in most of the patients (93%, 93/100 patients). The HCV genotype 2 and 4 were present in two patients respectively and HCV genotype 3 in one patient. Dual infection with genotype 1 and 3 was also identified in two patients. Sustained viral response was achieved in 56% (56/100) of the treated patients, confirmed by RT-PCR assay with detection limit of 20 IU/ml (Table 1).

The distribution of the frequencies of rs12979860 genotypes in the study group was as follows: 36 (36%) patients with CC genotype, 36 (36%) with CG genotype, and 28 (28%) with GG genotype. The sustained viral response rate in all study participants.

Table 1: The demographic features, distribution of HCV genotypes and the treatment response rate in all study participants.

<table>
<thead>
<tr>
<th>Patients, Number</th>
<th>100</th>
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<tbody>
<tr>
<td>Gender, Number (%)</td>
<td></td>
</tr>
<tr>
<td>Man</td>
<td>71 (71.0%)</td>
</tr>
<tr>
<td>Women</td>
<td>29 (29.0%)</td>
</tr>
<tr>
<td>Age, year, mean ± SD</td>
<td>43.5 ± 11.4</td>
</tr>
<tr>
<td>HCV genotype, Number (%)</td>
<td></td>
</tr>
<tr>
<td>G1</td>
<td>93 (93.0%)</td>
</tr>
<tr>
<td>G2</td>
<td>2 (2.0%)</td>
</tr>
<tr>
<td>G3</td>
<td>1 (1.0%)</td>
</tr>
<tr>
<td>G4</td>
<td>2 (2.0%)</td>
</tr>
<tr>
<td>G1 + G3</td>
<td>2 (2.0%)</td>
</tr>
<tr>
<td>Sustained viral response, Number (%)</td>
<td>56 (56.0%)</td>
</tr>
</tbody>
</table>

The achieved SVR in the study was 56% and it was associated with our study, to evaluate their association with the treatment-induced clearance of HCV in the patients treated with PEGIFN/RBV. The genome wide association studies identified that the homozygosis for the C allele of rs12979860, the homozygosis for the A allele of rs12980275 and the homozygosis for the T allele of rs8099917 were favorable genotypes which predicted treatment response. The same favorable genotypes were associated with the achievement of sustained viral response in treated patients in our study. Most of the studies investigated the effect of the IL28B gene polymorphisms on the SVR in patients with HCV genotype 1. Almost all of treated patients in our study were infected with HCV genotype 1, only 7% of the patients were infected with genotypes 2, 3 and 4. The study of Alam et al. [15], confirmed similar effects of the IL28B gene polymorphisms on the SVR in Caucasian patients infected with HCV genotypes 2, 3 and 4 as in the patients with HCV genotype 1.

The distribution of frequencies of rs12979860 genotypes in our study group was: CC in 36%, CT in 51% and TT in 13% of the participants. In Latvia, majority of the people are Caucasians, and they reported similar distribution of frequencies of rs12979860: CC in 33%, CT in 53% and TT in 14% of study participants (Caucasians, n=142) [16]. Similar to our findings, the distribution of frequencies of rs8099917 genotypes among study participants (Caucasians, n=175) in the study of Sticchi et al., was as follows: TT in 55%, TG in 40% and GG in 5% [17].

The achieved SVR in the study was 56% and it was associated with
the favorable genotypes of the IL28B gene polymorphisms. Consistent to our findings were the results from the study of Domagalski K et al. [18]. The predictability of the three most widely studied SNPs on the treatment response was determined in the cohort of 174 Caucasian (Polish) patients infected with HCV genotype 1 and 4 treated with PEGIFN/RBV. The CC genotype of rs12979860, TT genotype of rs8099917, and AA genotype of rs12980275 were significantly associated with the successful treatment (p=0.001, p=0.016, and p=0.002, respectively) [18]. Similar SVR rates and association with the IL28B gene polymorphisms were evaluated in the studies of Sporea I et al. [19] and Tolmane I et al. [16], although they studied only the rs12979860 as SNP. The study of Sporea I et al. included a cohort of 107 Caucasian (Romanian) patients infected with HCV genotype 1 treated with PEGIFN/RBV [19]. The SVR rate was 50.5% and there was a significant association between the SVR and CC genotype of rs12979860 (73.1% in the CC genotype vs. 43.7% in the non CC genotypes, p=0.012). The study of Tolmane I et al. included 142 Caucasian (Latvian) patients infected with HCV genotype 1 (61%) and HCV genotype 2 or 3 (39%) treated with PEGIFN/RBV [16]. The SVR rate was 59%, and it was significantly associated with the CC genotype of rs12979860 (74% in the CC genotype vs. 52% in the non CC genotypes, p=0.002).

Conclusions

The IL28B gene polymorphisms are strong predictors of the responsiveness to interferon-based regimen, and their determination has demonstrated a great potential to improve patient care. To date, direct antiviral agents including first and second generation protease inhibitors and the polymerase inhibitors represent the available options for highly effective interferon containing and interferon-free regimens for HCV eradication. In near future, even more direct antiviral agents will be approved for various combinations of interferon-free treatment options. However, in many countries approval and reimbursement of direct antiviral agents will be delayed. Also, PEGIFN/RBV therapy may be cost effective option to achieve an SVR in a subgroup of patients with beneficial genetic predictors of treatment response. Additionally, patients who have contraindications to these newer therapies will still need an interferon-based regimen, and thus the IL28B gene polymorphisms will still be important in predicting treatment response and prognosis.

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References