

Research Article

Dissolution testing of prolonged-release tablets using experimental design approach

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Abstract

Dissolution testing is an essential tool in the pharmaceutical industry and is used in formulation and process development, in monitoring of the manufacturing process, as a quality control test, to predict the in vivo performance. The purpose of this study is to evaluate the behaviour of the test and reference products using conventional dissolution apparatus basket and paddle, with tendency to evaluate the dependence of the in vitro dissolution of the dissolution test conditions (dissolution apparatus, medium, agitation, pH). Experimental design (ED) approach has been employed for assessment of the discriminatory properties of different dissolution conditions. The responses statistically evaluated were: f_2 similarity factor and the difference in the dissolution rate between the test and reference product, expressed in percentage, at every time point. Furthermore, this study focusses on developing a statistically reliable mathematical model for predicting discriminatory experimental conditions.

Introduction

A logical, systematic approach taking into consideration both scientific and regulatory principles, should be followed when developing a dissolution method. Properly designed dissolution tests will guide and accelerate drug development, assess batch to batch quality of a drug product, compare new or generic formulations with an existing product, assess the stability of the drug product, ensure the product quality in case of certain scale-up and post approval changes and provide a basis for achieving an *in vitro/in vivo* correlation. The method should also be challenged to discriminate between batches of material with different quality attributes.

Recently, Experimental Design (ED) have been widely used to understand the effects of multidimensional and interactions of input factors on the output responses of pharmaceutical products and analytical methods [1]. ED is a systematic approach to dissolution method development that begins with predefined objectives, emphasizes product and process understanding and sets up process control based on sound science. The knowledge obtained during development may

be used to support the establishment of an operable design space with suitable process controls during the life cycle of the product. These principles are now being considered for application to analytical methods through what is being termed Analytical Quality by Design (AQbD) [2].

In this study, the experimental condition parameters for dissolution testing of the drug product have been evaluated. In order to characterize the release from the dosage form adequately, it is recognized that a drug release profile should be generated, in which release (dissolution) values are determined as a function of time [3]. The experimental test conditions should be discriminating enough (“mild” conditions) to detect formulation and manufacturing variables that may affect pharmaceutical product performance. The following independent variables have been considered: media volume, different pH media, use of apparatus (basket and paddle) and different rotation speed. The purpose of this dissolution study is to evaluate the behaviour of the test and reference products to determine the dependency of the in vitro dissolution of the dissolution test conditions [4]. The objective is to assess the discriminatory potential of the dissolution method dependent



from all the regulatory permitted parameters. The responses statistically evaluated are: f_2 similarity factor and the difference in the dissolution rate between the test and reference product, expressed in percentage, at every time point.

Experimental

Materials and methods

Chemicals and reagents: Analytical grade acetonitrile, methanol, sodium heptansulfonate, phosphoric acid, sodium hydroxide, sodium chloride, hydrochloric acid, sodium acetate, acetic acid and potassium hydrogen phosphate were purchased from Merck (Darmstadt, Germany). Water was purified by a Werner water purification system, obtained in-house at Alkaloid AD Skopje, Skopje, Republic of North Macedonia.

Dissolution method: Dissolution testing was performed using 708-DS Dissolution Apparatus. Media used were: Simulated gastric fluid without enzymes (SGF), pH 4.5 acetic and pH 6.8 phosphoric prepared according to current Ph. Eur. requirements (5.17.1 Recommendations on dissolution testing). The temperature of the dissolution medium maintained at 37 ± 0.5 °C. In all experiments, the evaluated time points are the following: 15', 30', 60', 90', 2h, 4h, 6h, 8h, 10h and 12h.

Experimental design: Experimental runs were designed using software MODDE Go ver.12.0.1.3948 (Umetrics, Sweden). D-optimal quadratic design was applied for examining 4 independent variables (factors) at different levels within 24 runs. All the parameters from the dissolution method have been included: media volume, different pH media, use of apparatus (basket and paddle) and different rotation speed. The levels of the independent variables include the entire working spectrum from regulatory aspect. Summary of the design, the coded and actual experimental levels based on two-factorial three-level approach and the experimental matrix are given in Tables 1–3 respectively. The selection of the input factors was performed with intention for the design space to include all the regulatory permitted dissolution parameters. In order to gain more information, the design was enabled with “centre points” option to include mid values of each factor in the experimental design to check for lack of fit and curvature. Thus, a total of 24 runs were arrived for experimentation including 3 centre points.

Chromatographic method: Quantification was performed using HPLC method. HPLC studies were carried out on Agilent 1200 Series liquid chromatograph (Agilent Technologies Inc., Santa Carla, USA) equipped with a Photo diode array detector and UV detector. The mobile phase was comprised of 1.1 g/l sodium heptansulfonate solution adjusted to pH 2.0, acetonitrile and methanol in ratio 70:10:20 (v/v/v) at a flow rate of 0.8 ml/min. Kromasil C18 150 mm x 4.6 mm i.d; 5 μ m column was used, maintained at 55°C. Detection was at 230 nm wavelength. Run time of 12 minutes is utilized with injection volume of 50 μ l from the sample and the standard solution. Standard solution was prepared in final concentration of 0.08 mg/ml. Dissolution sample solutions at predefined intervals were withdrawn and filtered through 0.45 μ m regenerated cellulose membrane filter.

Results and discussion

In order to predict the variability of the results for response variables in relationship with the independent variables within proposed acceptance criteria, the results from the experiments were evaluated. Partial least squares technique (PLS) has been used as a nonlinear regression method for fitting a model to the data. This type of analysis has two objectives: to approximate the response variables and independent variables

Table 1: Summary of design.

Objective	Optimization (RSM)
Process model	Quadratic
Design	D-optimal
Runs in design	21
Center points	3
Replicates	0
N = actual runs	24

Table 2: Independent variables used for optimization study.

Independent variable	Type of factor	Levels
Apparatus	Qualitative	Basket, paddle
Dissolution media	Qualitative	SGF; pH 4.5; pH 6.8
Rotation speed	Multilevel	50; 75; 100 rpm
Volume	Multilevel	300; 500; 900 ml

Table 3: Experimental matrix.

Exp No	Exp Name	Apparatus	Medium	Rotation speed	Volume
1	N1	paddle	SGF	50	300
2	N2	basket	pH 4.5	50	300
3	N3	paddle	pH 6.8	50	300
4	N4	basket	SGF	75	300
5	N5	paddle	pH 4.5	75	300
6	N6	paddle	SGF	100	300
7	N7	paddle	pH 4.5	100	300
8	N8	basket	pH 6.8	100	300
9	N9	paddle	pH 4.5	50	500
10	N10	basket	pH 6.8	50	500
11	N11	basket	pH 4.5	100	500
12	N12	paddle	pH 6.8	100	500
13	N13	basket	SGF	50	900
14	N14	paddle	pH 4.5	50	900
15	N15	basket	pH 6.8	50	900
16	N16	paddle	pH 6.8	50	900
17	N17	paddle	SGF	75	900
18	N18	basket	pH 4.5	75	900
19	N19	basket	SGF	100	900
20	N20	paddle	pH 4.5	100	900
21	N21	basket	pH 6.8	100	900
22	N22	paddle	pH 6.8	75	500
23	N23	paddle	pH 6.8	75	500
24	N24	paddle	pH 6.8	75	500



and to maximize the correlation between them in the projected space. Quadratic mathematical model that best describes the relationship for each of the output factors.

Evaluation of the validity of the model was performed with ANOVA lack of fit (Figure 1). The lack of fit plot compares the Lack of Fit (LoF) component to the pure error component and displays a graph with 3 bars. The critical F is the value of the F-distribution over which SD LoF is statistically significant at the 95% confidence level. Since the first bar is smaller than the

third, the lack of fit is not significant at the 5% level, thus the model fits the data.

Adequacy of the used model was evaluated using summary of fit. Summary of the basic model statistics for four indicators R²; Q²; model validity and reproducibility are presented for all the response variables (Figure 2). R² shows the model fit. The values (0.70–0.92) are close to 1 and indicate that the model fits the data very closely. Q² represents an estimate of the future prediction precision. Model validity is a measure of the

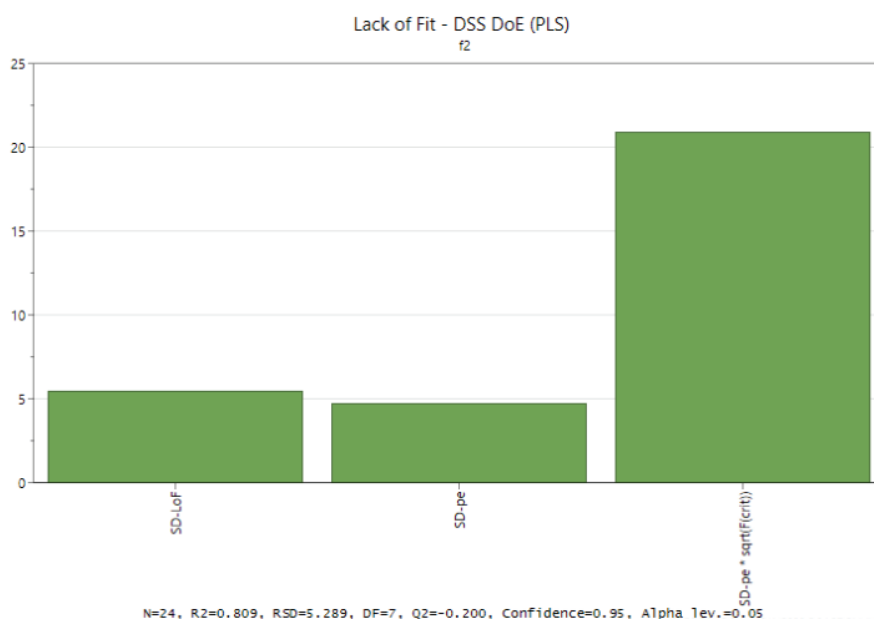


Figure 1: Lack of fit. The lack of fit plot compares the Lack of Fit (LoF) component to the pure error component and displays a graph with 3 bars. SD-LoF: Shows the variation of the response due to the lack of fit of the model (i.e. the model error) adjusted for degrees of freedom and in the same units as Y. This is the square root of MS (mean square) lack of fit. SD-pe (Pure error): Shows the variation due to the replicated experiments (observations) adjusted for degrees of freedom and in the same units as Y. This is the square root of MS (mean square) pure error. SD-pe*sqrt(F(crit)): Shows SD pure error (second bar) multiplied by the square root of the critical F.

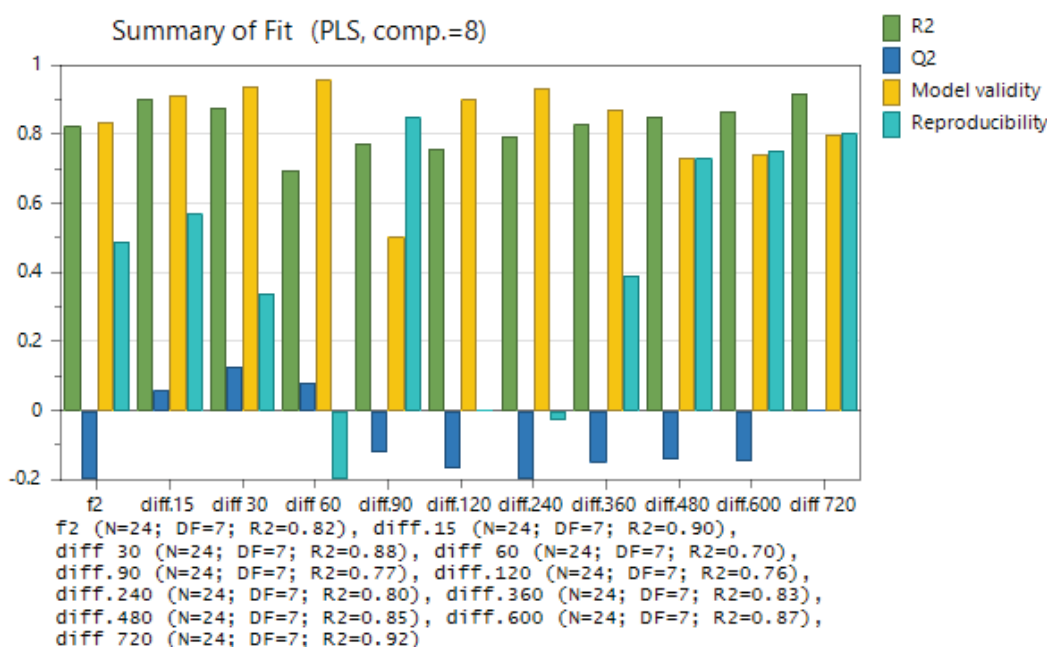


Figure 2: Summary of fit (Partial least squares regression-PLS).



validity of the model and a test of diverse model problems. The model validity column (>0.5) is larger than 0.25, hence there is no Lack of Fit of the model (the model error is in the same range as the pure error). Reproducibility presents the variation of the replicates compared to overall variability. Summary statistics for these indicators in relation to concerned response variables: apparatus, medium, rotation speed and volume shows that the model is significant and fits the data, since the model performance indicators comply with the above presented reference values.

The qualitative contribution of each factor on the responses were analysed by Pareto chart (Figure 3). Pareto charts establish critical value of the effect. Coefficients with value of effect above 1 are designated as certainly significant. In this

case all investigated effects are considered as insignificant. Nevertheless, most influential input factors on the similarity factor response are the rotations per minute and the volume of the media.

Evaluating output functions, it was taken into consideration that the guidelines have requirements for assessment of similarity of the dissolution profiles. Since none of the factors is statistically significant for the output factors - differences in different time points, the interactions have been evaluated only for f_2 as a response (Table 4). Interactions are evident only between apparatus type and volume. The coefficients for this interactions are 0.2096 and -0.2096 for apparatus (basket)*volume and apparatus (paddle)*volume, respectively. The apparatus is a qualitative factor, which means that the

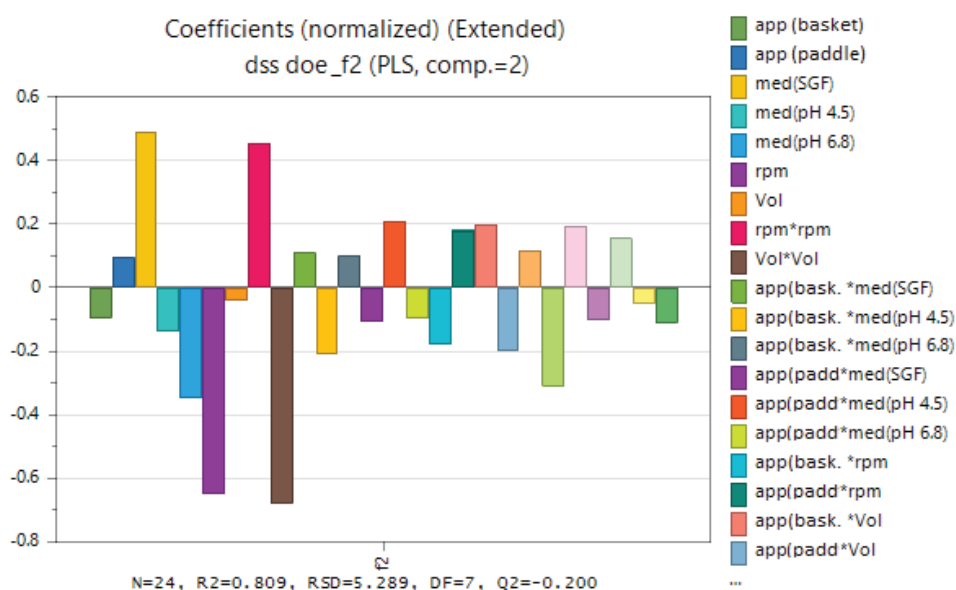


Figure 3: Pareto chart showing the effects based on the observation of D-optimal design.

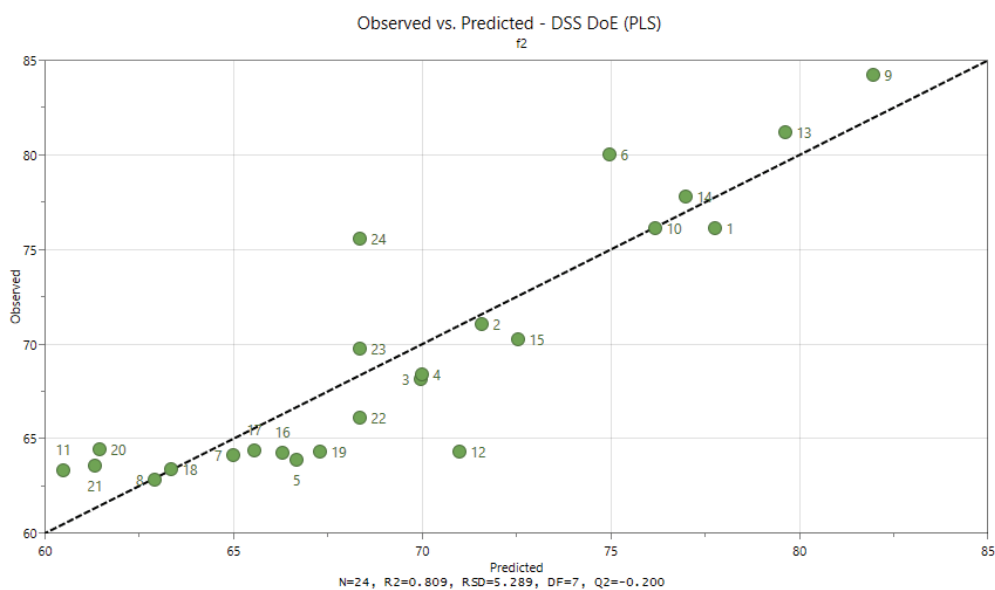


Figure 4: Observed vs. Predicted plot.



value of the coefficients origins from the influence of the volume, increasing the volume contributes to increasement of the similarity factor f_2 . Furthermore, the volume has different effect on different type of apparatus.

Additionally, observed versus predicted values have been

evaluated (Figure 4). Plot with points close to a straight line indicate good model.

The Response Contour Plot displays the predicted response variables values for the selected response variables, spanned by two factors, in a response surface contour plot. It is used to determine where a maximum (red colour) or minimum (blue colour) response is expected taking in account the independent variables. The 4D response contour of f_2 is displayed with the response variables: media volume, apparatus basket or paddle, medium type and rotation speed (Figure 5).

Sweet Spot Plot highlights the areas where the responses are within specified ranges (Figure 6).

Response variables are given in Table 5. An f_2 value between 50 and 100 suggests that the two dissolution profiles are

Table 4: Interactions between input factors.

Type of interaction	Interaction present
apparatus * medium	-
apparatus * rotations per minute	-
apparatus * volume	+
medium * rotations per minute	-
medium * volume	-
rotations per minute * volume	-

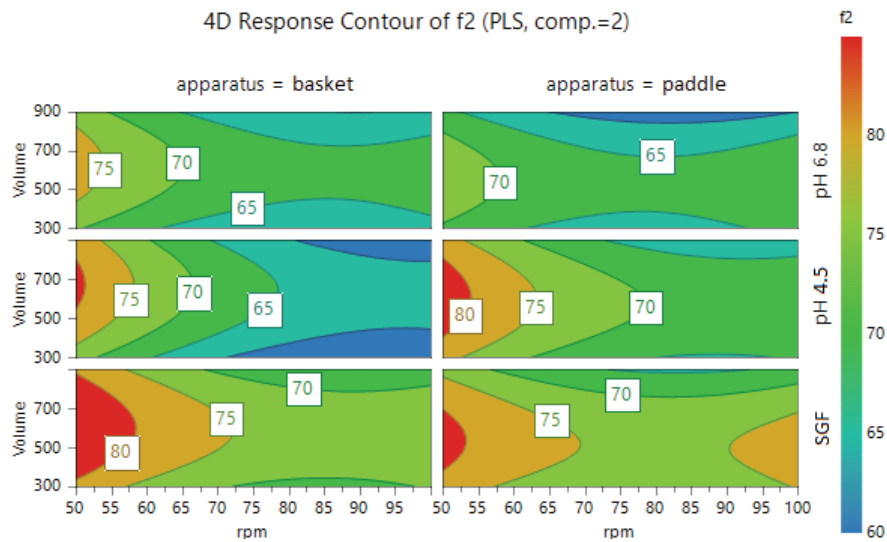


Figure 5: 4D Response Contour Plot of f_2 (PLS, comp.=2).

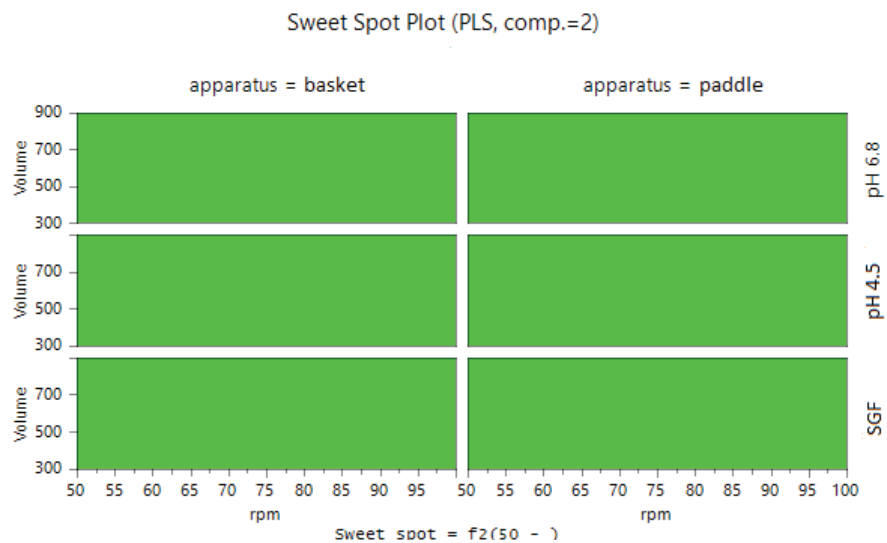


Figure 6: Sweet spot plot. The plot uses the colour scale from green to blue with: Green for the "sweet spot", that is the area where all the responses are within the selected range; Blue for the areas where none of the responses are within their selected ranges; Other colours for areas with more than one response within its range but not all.

**Table 5:** Response variables used for optimization study.

Exp. No	Exp. Name	f_2	diff. 15'	diff. 30'	diff. 60'	diff. 90'	diff. 120'	diff. 240'	diff. 360'	diff. 480'	diff. 600'	diff. 720'
1	N1	76.14	0.79	1.20	1.22	1.46	0.75	2.64	6.54	8.63	7.82	6.49
2	N2	71.08	4.41	4.02	1.22	0.12	1.28	3.95	6.25	7.98	8.12	5.87
3	N3	68.17	0.41	0.48	0.19	1.27	2.32	6.06	8.95	9.68	8.78	7.37
4	N4	68.39	4.76	4.41	1.69	0.33	0.75	4.03	7.73	10.38	10.00	6.86
5	N5	63.88	1.59	2.03	3.51	4.38	5.62	7.20	8.14	7.21	7.36	4.55
6	N6	80.05	0.03	0.54	0.36	0.29	0.12	3.11	5.17	5.69	2.34	4.68
7	N7	64.15	1.77	1.72	2.70	4.94	7.17	6.25	7.41	10.53	6.99	3.73
8	N8	62.84	3.29	3.89	0.66	1.58	3.59	7.39	10.54	12.65	12.42	10.42
9	N9	84.22	1.49	1.89	2.56	0.34	1.58	0.45	2.21	2.48	1.32	0.39
10	N10	76.14	0.99	1.16	0.40	0.33	1.11	3.11	4.89	5.16	4.15	2.18
11	N11	63.31	1.33	1.58	0.16	1.27	2.27	5.56	12.51	12.23	7.26	3.17
12	N12	64.34	1.23	0.96	1.15	2.42	3.33	7.84	9.88	9.94	9.66	8.80
13	N13	81.23	0.66	1.05	0.35	0.09	0.51	2.61	4.87	5.73	4.50	2.12
14	N14	77.77	1.17	1.43	1.67	1.94	0.71	0.59	6.03	6.82	4.96	3.18
15	N15	70.25	0.79	1.06	0.38	1.27	3.46	5.13	7.72	7.97	4.59	3.00
16	N16	64.25	0.44	0.06	0.46	1.88	2.99	7.87	10.32	11.04	10.22	8.48
17	N17	64.38	0.92	0.89	0.38	1.78	3.39	5.80	9.02	8.56	5.51	4.97
18	N18	63.41	0.41	1.09	2.40	3.33	4.30	7.24	10.37	12.55	10.32	6.75
19	N19	64.32	0.45	0.77	1.45	2.51	3.42	6.30	10.94	14.86	9.83	6.39
20	N20	64.43	0.61	0.12	1.73	2.87	4.38	7.43	9.60	14.56	10.45	7.98
21	N21	63.55	0.21	0.42	0.86	2.91	4.09	7.62	10.47	11.53	10.43	11.17
22	N22	66.09	2.26	2.17	2.41	1.40	3.37	6.83	8.75	8.85	7.70	6.20
23	N23	69.78	0.15	0.02	0.03	0.71	1.71	6.02	8.15	8.20	7.72	5.79
24	N24	75.55	0.27	0.06	0.59	0.53	0.05	2.47	5.22	5.83	5.12	3.78

diff. - difference in the dissolution rate between the test and reference product, expressed in percentage, at every time point

similar³. The f_2 similarity value is between 62.84 and 84.22. The difference of the percent released in the time points up to 2 hours is less than 5%, which confirms the similarity of the dissolution profiles of both products.

Conclusions

The goal of a well-characterized method development effort is to characterize and develop a reliable method that can be demonstrated with a high degree of assurance to consistently produce data meeting predefined criteria when operated within defined boundaries.

From the above data it can be concluded that all the variable conditions of the dissolution testing with conventional apparatus basket and paddle result in similar dissolution profiles of the test and reference product. According to the sweet spot plot, design space of the analytical method can be defined where the results are independent from the test conditions. *In*

vitro dissolution of both drug products is independent of the dissolution test conditions. The proposed experimental design approach was able to identify critical factors and the gained information could aid in general decision making.

References

1. Fakuda IM, Pinto CFF, Moreira CS, Saviano AM, Lourenco FR (2018) Design of Experiments (DoE) applied to Pharmaceutical and Analytical Quality by Design (QbD). 54: e01006. [Link: https://bit.ly/2GAgm1j](https://bit.ly/2GAgm1j)
2. Borman P, Nethercote P, Chatfield M, Thompson D, Truman K (2007) The application of quality by design to analytical methods, *Pharmaceutical Technology* 31: 142-152. [Link: https://bit.ly/3ixHueM](https://bit.ly/3ixHueM)
3. EMA (2010) Guideline on the Investigation of Bioequivalence. CPMP/EWP/QWP/1401/98 Rev.1/Corr**. [Link: https://bit.ly/3ixHCuM](https://bit.ly/3ixHCuM)
4. FDA, CDER (1997) Guidance for Industry: Extended Release Oral Dosage Forms: Development, Evaluation, and Application of In Vitro/In Vivo Correlations. [Link: https://bit.ly/3jzVDJF](https://bit.ly/3jzVDJF)